Parameter estimation : introduction
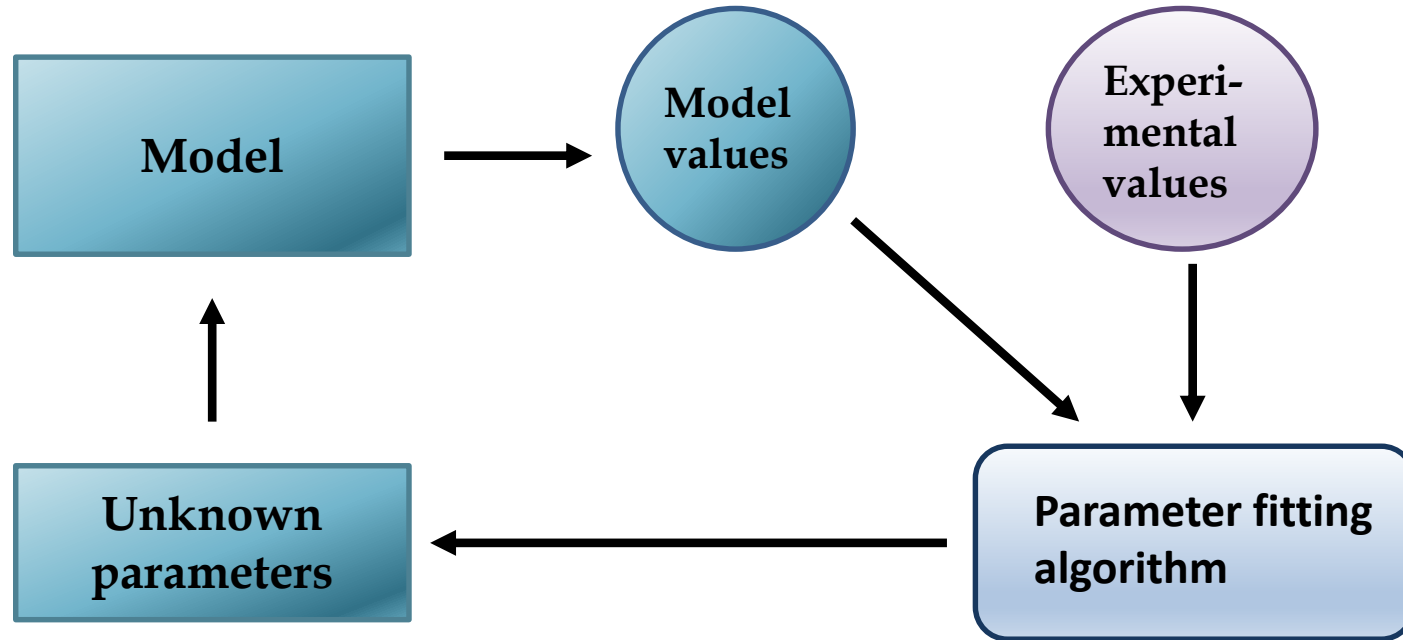
When starting a modeling project usually many parameters of the model are not known

How can I find out about parameter values?

➤ experimental approach: try to design an experiment for measuring the specific parameter

- typically *in vitro* experiment (e.g. for rate constants: put different amounts of substrate in a test tube and measure how fast the reaction proceeds)
- Problems: often not possible, to many parameters

➤ Systems biology approach: adapt a complete model to experimental data

Parameter estimation : basic idea of parameter fitting

Change the parameter values of a model in order to that it best fits the experimental data

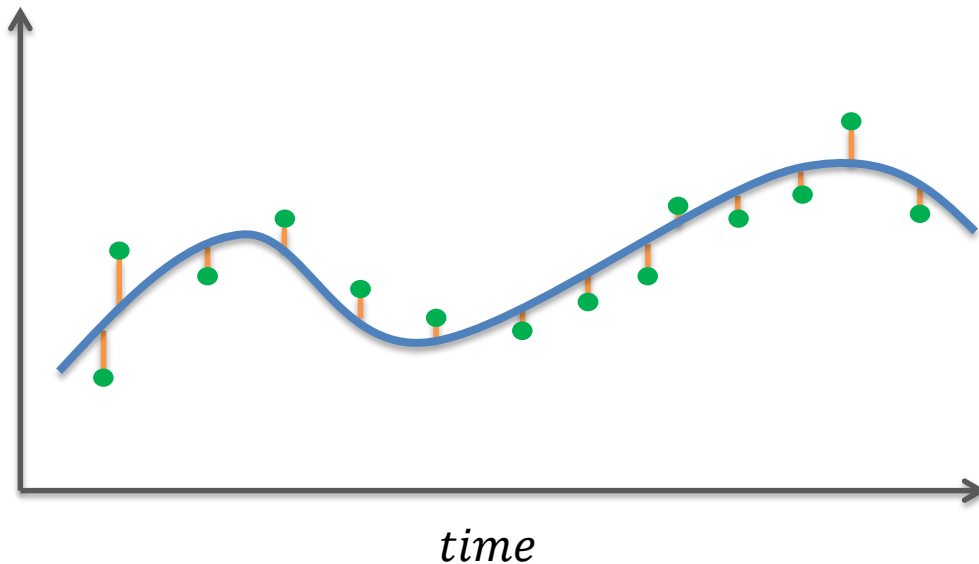# Parameter estimation : basic idea of parameter fitting

How to determine the "best" fit for a given set of experimental data ?

We will use heuristics

- Computation of the probability that the measurements (experimental data) would be the results of a simulation of the model

- A high probability means that the model is good

- Criterion: likelihood is maximal when the difference between measurement and simulation results is minimal (easy to calculate)
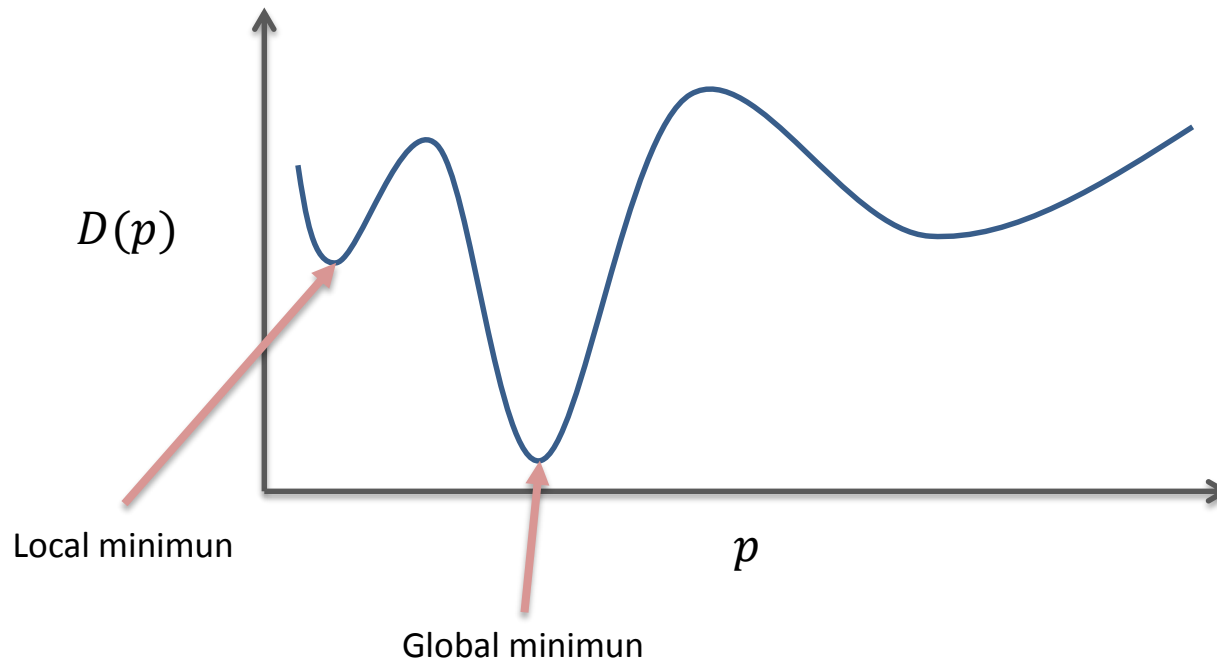
Parameter estimation

The parameters inference problem for ordinary differential equation models is usually formulated as an optimization problem with an objective function that has to be minimized by adjusting the values of the model parameters. A common choice to compute this objective function is to calculate the sum of squared errors between measurements and model predictions : the least squares distance measure
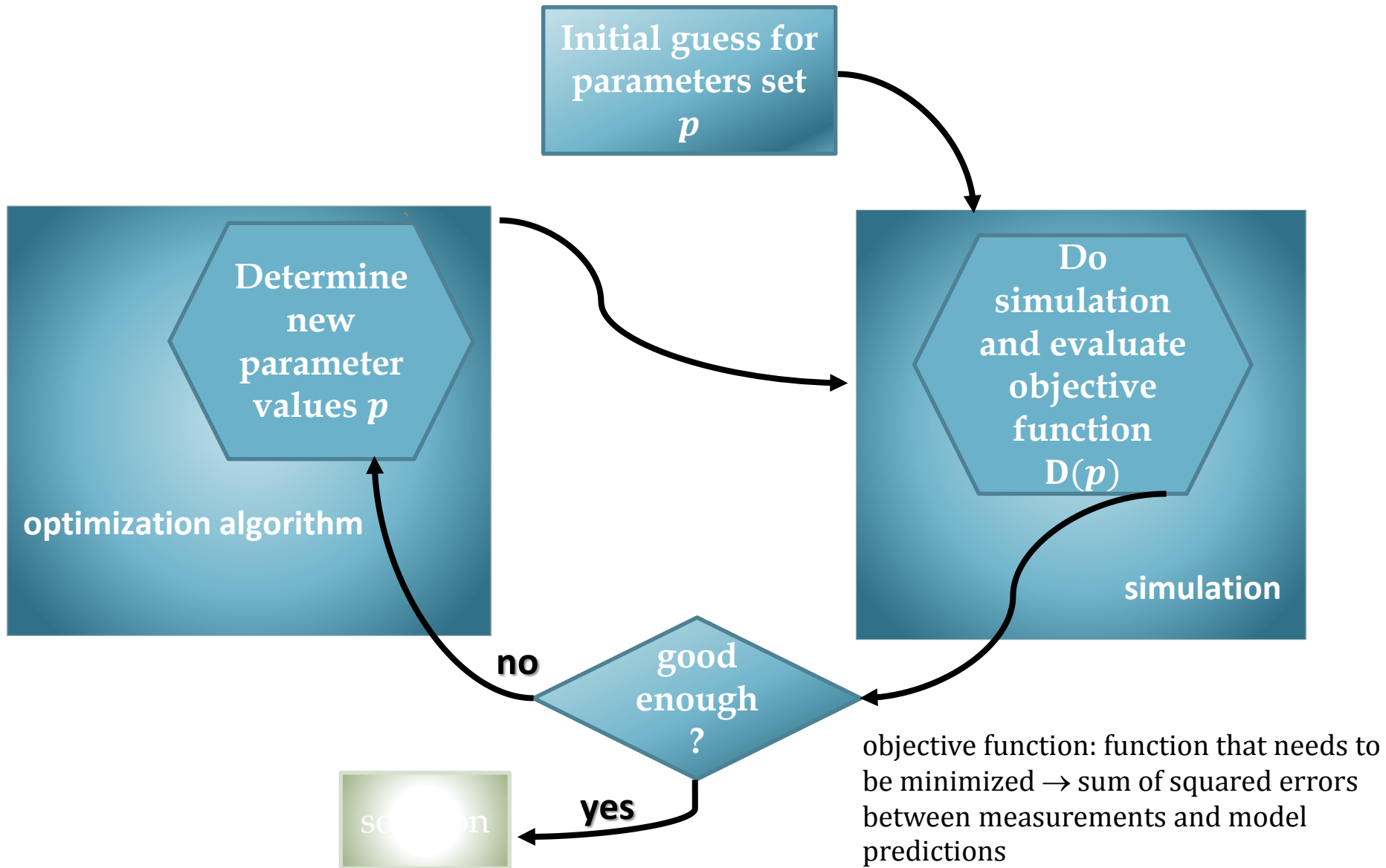
$$D(p) = \sum_{i=1}^{N}(x_i - y_i(p))^2$$

A compound is tracked over time and we obtained N values. Thus N is the number of data point, x_i is the measured value for time t_i and , $y_i(p)$ is the simulated value for time t_i for a set of parameter values p

time

Parameter estimation : parameter space

$D(p)$

Local minimun

Global minimun

p

- The objective function may have many local minima
- If we have *N* parameters to estimate, the parameters represent an *N*-dimensional space, the parameter space
- A specific solution (specific parameter values) will correspond to a point in the parameter space
- We need a way to find the set of parameter values (a point in parameter space) for which the distance D is minimal (the best fit)
- We will use an optimization algorithm since a systematic scan of the parameter space is not possible when the space dimension is large

Parameter estimation : numerical optimization cycle

objective function: function that needs to be minimized → sum of squared errors between measurements and model predictions

Parameter estimation : optimization algorithm

Different methods have been proposed:
Local optimization methods tend to converge quickly but have a tendency to get stuck in local optima
Global methods might take time but ensure the global optimum

Focus on the particle swarm optimization (PSO) method since its performance compared to the fourth most popular optimization methods (Evolutionary Computation, Evolutionary Programming, Genetic Algorithms and Simulated Annealing) reveals that the PSO method performs the best in systems biology (Baker *et al*, 2010, Journal of Integrative Bioinformatics, 7(3):133).

PSO belongs to the class of stochastic global optimization methods which depend on probabilistic approaches.

PSO (optimization par essaims particulaires) was first proposed by Kennedy and Eberhart (1995, In *Proceedings of the Sixth International Symposium on Micro Machine and Human Science MHS95, IEEE Press*, 3943). It is inspired by social behavior and movement dynamics of insects, birds and fishes.

Parameter estimation : particle swarm optimization method

The swarm is typically modeled by particles that have a position and a velocity in multidimensional space. These particles roam through the hyperspace and have two essential reasoning capabilities: A particle is described by a position vector in the parameter space and a velocity vector. The velocity of a particle represents the direction of its parameter space exploration and the speed of the movement.

Each particle possesses:
•   the memory of its own previous experience and remembers its best achievement *(pbest)* Information about the best solution attained within its neighbors . The "global" version of the optimizer keeps track of the overall best value, and its location, obtained thus far by any particle in the population (*gbest*). In the "local" version, each particle keeps track of the best solution attained within a *local* topological neighborhood of particles (*lbest*)

Each particle of the swarm is randomly initialized for its position and velocity. These particles roam through the parameter space and optimization concept consists of, at each time step, changing the velocity (accelerating) of each particle toward its *pbest* and *gbest or lbest.*

Here, a particle will correspond to the set of kinetic parameters that have to be estimated and a best achievement corresponds to the smallest value obtained for the objective function

Parameter estimation : particle swarm optimization method

Let S be the number of particles in the swarm.
Each particle i has a position $\boldsymbol{x}_i \in \mathbb{R}^n$ in the search-space and a velocity $\boldsymbol{v}_i \in \mathbb{R}^n$
Let $\boldsymbol{p}_i$ the best known position of particle i and $\boldsymbol{n}_i$ the best known position of its neighbor
Let f the objective function which must be minimized

Initialization
    for each particle i = 1, ..., S do
        Initialize randomly the particle position vector $\boldsymbol{x}_i$
        Initialize the particle's best position $\boldsymbol{p}_i$ with $\boldsymbol{x}_i$ ($\boldsymbol{p}_i \leftarrow \boldsymbol{x}_i$)
        If $f(\boldsymbol{p}_i) < f(\boldsymbol{n}_i)$ then
        Update the neighbor's best position $\boldsymbol{n}_i \leftarrow \boldsymbol{p}_i$
        Initialize the particle velocity

While a termination criterion is not reached do
    for each particle i = 1, ..., S do
        Update the particle velocity as follow:
        $\boldsymbol{v}_i = w\,\boldsymbol{v}_i + c_1\,r_1\,(\boldsymbol{p}_i - \boldsymbol{x}_i) + c_2\,r_2\,(\boldsymbol{n}_i - \boldsymbol{x}_i)$
        Update the particle position:
        $\boldsymbol{x}_i = \boldsymbol{x}_i - \boldsymbol{v}_i$
        If $f(\boldsymbol{x}_i) < f(\boldsymbol{p}_i)$ then
            Update the particle best known position $\boldsymbol{p}_i \leftarrow \boldsymbol{x}_i$
            If $f(\boldsymbol{p}_i) < f(\boldsymbol{n}_i)$ then
                Update the neighbor's best position $\boldsymbol{n}_i \leftarrow \boldsymbol{p}_i$

termination criterion:
the specified number of iteration or the value
of the objective function is < than a given
threshold and a solution has been found

w = inertia
r_1 and r_2 = random number
In general $c_1 = c_2 = 2$

Parameter estimation : particle swarm optimization method

Each iteration in PSO execution, requires for each particle of the swarm:

1. the values of the position vectors are set as the values of the model parameters
2. the model is simulated, by numerical integration of the ODE system, to produce the dynamic profiles corresponding to those parameter values
3. the simulated data are compared to the experimental data using the objective function described above

The iterative process will stop if the change of the objective function value is smaller than a specified value or if the number of specified iterations is reached.

Parameter estimation

Important:

• The result of a parameter fitting always needs to be inspected afterwards

• Having a good result for a fit does **not** mean that the parameter value is the „true" one. This depends on the assumptions about the errors and the correctness of the model.

• For the stochastic algorithms the result is not reproducible

Parameter estimation in COPASI

Experimental data

Example of experimental time-series data: transcriptional fusion of a fluorescence (GFP) or luminescence (luciferase) reporter gene to the promoters of the target genes

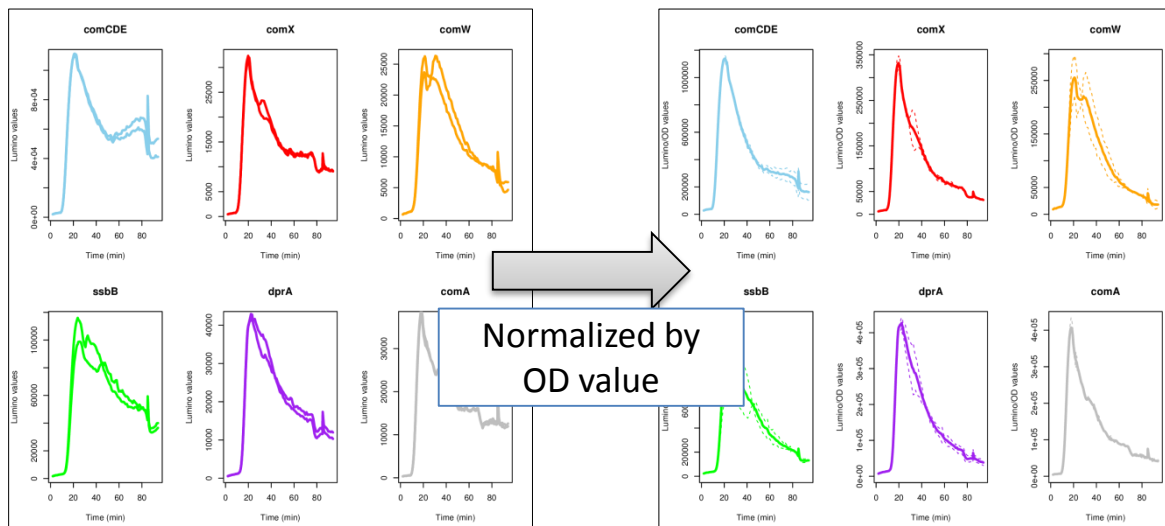Measurement techniques allow real-time and in-vivo monitoring of gene expression

Zulkower *et al., Bioinformatics* 2015

Ronen *et al., PNAS* 2002

Normalization of luminescence data

The quantity of luminescence per cell as a function of time ($r(t)$) is : $r(t) = \dfrac{I(t)}{A(t)}$ with

- $I(t)$ is the luminescence intensity (in RLU)
- $A(t)$ the absorbance values corrected by subtraction of the absorbance background measured on wells containing only growth medium

Since the luciferase does not require any post-translational modification such as folding, this ratio estimates the average concentration of reporter protein per cell.
The dynamics of the system is conveniently described by the temporal evolution of the luciferase concentration.
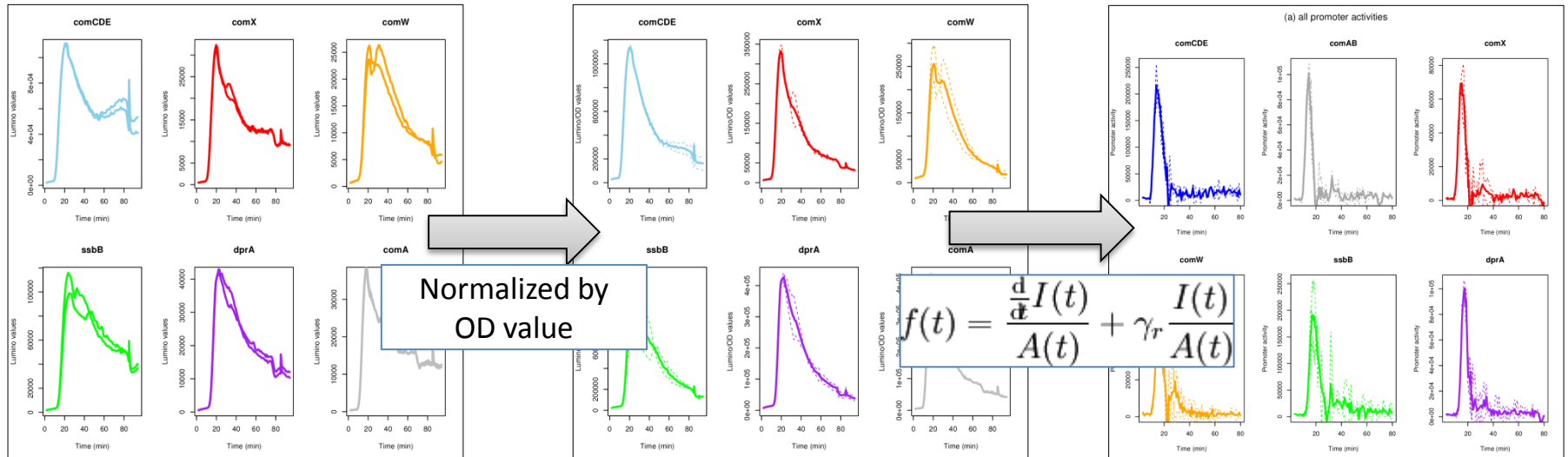
Use of the green fluorescent reporter gene

- Fluorescent activity of GFP in response to light excitation depends on post-translational modifications, notably the folding of the protein to an appropriate conformation, including the autocatalytic formation of the chromophore.
- This maturation process gives rise to an additional reaction step from GFP to active GFP.
- See de Jong *et al. BMC Systems Biology* 2010, **4**:55 for correct normalization of fluorescence signal

Promoter activities deduced from luminescence signal

Promoter activities are deduced using its derivative according to the following formula (Stefan *et al*, 2015, *PLoS Comput. Biol.* **11** e1004028):

$$f(t) = \frac{d}{dt} r(t) + \left(\gamma_r + \mu(t)\right) r(t) = \frac{\frac{d(I(t)}{dt}}{A(t)} + \gamma_r \frac{I(t)}{A(t)}$$

where γ_r [min^{-1}] is the degradation constant of the luciferase protein and $\mu(t)$ [min^{-1}] is the growth rate of the bacteria

protein concentration kinetics

Computation of the concentration evolution of the protein of interest over time:
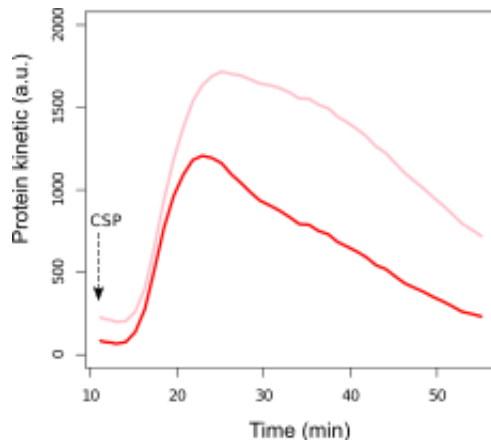→ Correction to take into account the differences in half-lives between the reporter luciferase and the protein whose gene activity is measured (Stefan *et al*, 2015, *PLoS Comput. Biol.* **11** e1004028):

$$\frac{d}{dt}p(t) = f(t) - \left(\gamma_p + \mu(t)\right)p(t), \qquad p(0) = p_0 = \frac{\mu(T) + \gamma_r}{\mu(T) + \gamma_p}\, r(T)$$

where γ_p [min^{-1}] is the degradation constant of the protein and $\mu(T)$ is the growth rate of bacteria at the end of the preculture procedure (at time T)

Correction effects:
*   protein of interest with a shorter life times than luciferase: the uncorrected values clearly overestimate the protein concentration (red curves)
*   protein of interest with a higher life times than luciferase: the uncorrected values underestimate the protein concentration (blue curves)

*   Dark color lines protein concentration kinetics taking into account only protein life time
*   Light color lines protein kinetics after correction